



Control of Heavy Metals from Barley and Wheat Grains during Malting and Brewing

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Abstract

Among seven wheat, rye, and pea crops from a non-remediable polymetallic soil with 39.6 mg kg⁻¹ in Cd, barley ranking fourth in grain production experienced the least yield losses in grains (37% vs. 60-74% in the other seed crops). With 0.455 mg kg⁻¹ by dry weight in Cd vs. 1.20-3.13 mg kg⁻¹ in the reference seeds, barley grains even met the limit of 1.0 mg kg⁻¹ fresh weight set for livestock feed and human food supplements. Grains of barley and the wheat cvs. JB Asano (Cd, 1.55 mg kg⁻¹) and Kavalier (Cd, 0.054 mg kg⁻¹) were therefore used for the production of malt extract and beer to estimate consequences for their misuse in food processing. Heavy metals (HM) were monitored by Inductively Coupled Plasma Mass Spectrometry (ICP-MS) after microwave digestion. During micromalting of barley and wheat grains, 74% and 83%, respectively, of Cd, Cr, Cu, Mn, Pb, U, and Zn, followed by As and Ni with around 34%, were removed from the mash with the spent grains that retained the cell wall-bound Cd, Cu, Ni, and Zn in values critical for livestock feed. With Cd concentrations of 0.019 and 0.041 mg kg⁻¹, respectively, the remaining turbid sweet wort used in 1:5.5 dilution met the limit of 0.1 mg kg⁻¹ for solid food. Its fermentation with *Saccharomyces cerevisiae* precipitated another 79-98% Cd and 31-93% As, Cu, and Zn with their highest affinity to organic binding sites of residual tissue fragments and proteins but no further Cr, Mn, Ni, Pb, and U from solution. With the widest separation of organic-bound HM from the fraction of fermentable sugars, the resulting transparent barley and wheat beers concurred with the guidelines for drinking water and alcoholic beverages. It is argued that the exceptionally low grain acquisition of Cd, Ni,

As, and Pb in barley accessions and the further HM control during processing shall not be concealed. If barley production is lightly extended to cropland with HM contaminations beyond regional guidelines, the exclusive use in husbandry and bioethanol production must be ensured to preclude future ignorance of any soil hygiene measures.

Keywords: Brewers' Wort; Cadmium; Malt; Mashing; *Saccharomyces Cerevisiae*, Spent Grains; Trub; Wheat and Barley Beer

Introduction

The need of a growing human world population in staple food, energy, industrial raw materials, and more stock farming products is uncontested. Nevertheless, these pretensions conflict with the loss of quality cropland to urbanization, sealing, mining, salinity, and land erosion [1-4]. Sewage and husbandry sludges, industrial effluents used as irrigation water, and arsenic (As), cadmium (Cd), and uranium (U) traces in phosphate fertilizers enrich heavy metals (HM) in soil [5-7]. In tolerating depreciated cropland, barley accessions with their greater acceptance of problematic soils, climatic drawbacks, and generally lower rates of As, Cd, nickel (Ni), and lead (Pb) in the grains than other cereals [8-10] should thereby be the first choice.

With an annual world production of 136 to 160 Mio Mg and yields of 1800-6400 kg ha⁻¹ that are surpassed by values of 7600-11,000 kg ha⁻¹ in industrialized regions [11, 12], barley is ranking fourth behind the grain crops wheat, maize, and rice. In regions of high altitude, aridity, salinity, and other

unfavorable conditions, the crop may become the only staple food resource [13-16].

Barley grains are used in malt, beer, and bioethanol production and serve as feed in husbandry [17]. Some health benefits such as heart disease, diabetes, and cancer prevention, body weight control, and reductions in total and LDL cholesterol [18-20] and blood sugar [21] are linked with barley consume. Barley straw is used for bedding in husbandry but also as roughage for ruminants in combination with alfalfa [22], hay, 14-30% proteins, a mineral mix, and salt [23, 24].

Among seven experimental crops of wheat, rye, and pea from a polymetallic soil with 39.6 mg kg⁻¹ in Cd and 154, 205, 39, and 3200 mg kg⁻¹ in As, Cu, U, and Zn, respectively, barley experienced the least losses in grains (37% vs. 60-74%) and straw (0% vs. 52-67%) in comparison to crops from non-contaminated soils [8]. Heavy metal concentrations in the herbage of barley ranged within those of other seed crops and excluded its use as roughage supplement and bedding in husbandry by exceeding the permissible limit of 1.0 mg kg⁻¹ [25] with 4.65 mg kg⁻¹ in Cd [8]. HM concentrations in grains met those of other cereals, too, but were drastically lower in Cd and Ni. With 0.455 mg kg⁻¹ by dry weight (DW) in Cd versus 1.20-3.13 mg kg⁻¹ in the other seed crops, barley grains from soils of 39.6 mg kg⁻¹ in Cd even met the limit of 1.0 mg kg⁻¹ fresh weight (FW) set for livestock feed and human food supplements [8, 25, 26]. This is alarming in regard to the observation of soil and crop hygiene standards and has so far not yet been reported. Thereby, carelessly created perils for consumers of beer, malt amended food, and stock farming products by making the most of the legislative hygiene limits for As, Cd, and Pb [26] must be excluded. Contaminated and non-contaminated barley and wheat grains were therefore used for the production of malt extract and beer to estimate consequences for their misuse in food processing, but also for the possibility to separate the fermentable sugar fraction from HM contaminated tissues.

Subjacent to the two husks in dorsal and ventral position of the barley grain, the living cells of the outer and inner covering layers of pericarp and testa enclose the embryo and the dead cells of the starchy endosperm to stabilize seed dormancy by restricting ingress of oxygen. The endosperm itself is surrounded by three cell layers of the aleurone and separated from the embryo by the epithelial cells of the scutellum [27, 28]. Starch granules of the endosperm account for 60-65% by DW of the grain. Starch is composed of 17-24% n-amylose that forms chains of 60-2000 glucose residues in α -1,4 bonds. Amylopectin as the main component forms branched chains of glucose units in α -1,4 and α -1,6 bonds [27].

With the production of gibberellic acid (GA3) by the germinating embryo, the aleurone releases hemicellulases, proteolytic enzymes, amylases, and phosphatases for the 'modification' of the endosperm. In the major steps, glucanases break down its cell walls that primarily consist of β -D-glucan. Proteins encompassing the starch granules are

degraded by proteases to lower molecular weight proteins, peptides, and amino acids. β -Amylase derives yeast-fermentable maltose from both starch components. α -Amylase generates oligodextrin (six glucose units) and branched dextrans that are further converted to maltose and maltotriose by β -amylase in the exclusive cleavage of α -1,4 bonds. The remaining 20% of limit dextrans with α -1,6 bonds are broken down by dextrin-6- α -D-glucanohydrolase to dextrans accessible to β -amylase [27, 28].

Industrial protocols for the production of malts (malted grains) start with steeping barley and wheat in three successive batches of water at 12-18°C to raise the grain water content to 43-48% by total weight over 22-28 h, interrupted by phases of aeration, and accompanied by a DW loss of 0.8%. Supported by water spraying, grains germinate within six days at temperatures as low as 14-18°C to reduce the loss of biomass by respiration. The green malt of 160 kg per 100 kg of grains is then slowly dried by 'kilning' to 80 kg and a water content of 3-4% at temperatures of 50-77°C, designed to support the grain internal enzymatic catalyses of starch to fermentable sugars. After removing the sprouts of roots and shoots that are enriched in trace metals by factors 2-6x [29], malt grains are crushed and mashed in aqueous suspensions of 1:2.5 to 1:4 (w/v). The fermentable sugars of the resulting heated sweet wort are formed by maltase (at 35-40°C), saccharase (50°C), dextrin-6- α -D-glucanohydrolase (55-60°C), β -amylase (62-<70°C), and α -amylase (72-<75°C). Intermediate boiling half of the mash supports the dissolution of starch. The final mineral stock of the resulting beer may be given by the input sources of the original grains and the Al- and Cu-pesticide treated hop, reduced by the metal binding fractions of spent grains or draff, and the trub precipitate formed by fermenting yeasts [27, 29-31].

In adaptation to the procedures given above, barley and wheat grains obtained from contaminated and non-contaminated arable soils were processed to malt extract and fermented to beer with the goal to record the gradual removal of critical HM, especially of Cd and Zn, with spent grains and trubs from the fraction of fermentable sugars under consideration of the ethics of food quality.

Materials and Methods

Grain Crops

Of the recent grain crops cultivated on polymetallic soils with abating HM load, a two-rowed winter barley cv. tested in the 2015/2016 season was chosen. It has been compared with the contaminated grains of the winter wheat cultivar JB Asano, an A-quality crop denoted by a high 1000-seeds FW and an average raw protein content as described in detail [32]. The winter wheat cv. Kavalier grown on non-contaminated arable derived from mottled sandstone in 2016/2017 [33] was used for comparative micromalting.

Soils

With the local inclusion of aged polymetallic overburdened soil from uranium mining into non-contaminated cropland, a gradient of geologically related soils with diminishing HM concentrations had been formed in the late 1960s. The clay-loam soil was derived from Permian

limestone, mottled sandstone, and dolomite [34]. Its mineral content was confirmed by the random collection of six soil samples from the 20-cm plough layer. Soil (sieve 0.8 mm) from the HM hot spot (Table 1) of pH_{aq} 7.12 contained 3.53% in C_{org}. The winter wheat cv. Kavalier was obtained from an arable soil of pH_{aq} 6.85 and C_{org} 1.55% as derived from mottled sandstone.

Cu	Mn	Ni	Zn	As	Cd	Cr	Pb	U
Heavy metal hot spot of polymetallic soil								
171 ± 5	1932 ± 4	41.7 ± 1	3207 ± 104	144 ± 5	39.6 ± 0.8	22.5 ± 1.6	161 ± 4	33.3 ± 0.6
Non-contaminated arable soil on mottled sandstone								
8.4 ± 1.2	198 ± 21	6.49 ± 0.56	33.1 ± 2.7	3.62 ± 0.38	0.105 ± 0.008	12.9 ± 2.1	9.4 ± 0.9	0.547 ± 0.056

Table 1: Heavy metal concentrations (mg kg⁻¹ DW ± SD) of the arable soils used for cropping the current grain sources (n = 6).

Mineral Concentrations in Soils and Grains

Mineral concentrations were determined for duplicate aqua regia-extracted 5-g samples of soils. Duplicate samples of fresh gains milled for analyses (Pulverisette 14; Fritsch GmbH, Idar-Oberstein, Germany) were prepared from 10 g in FW each after careful washing with bideionized water. Samples of 0.3 g were microwave-digested in HNO₃ (4 mL) and H₂O₂ (1 mL) (Mars Xpress; CEM GmbH, Kamp-Lintfort, Germany) and diluted to 150 mL with bideionized water. The solutions were analyzed by Inductively Coupled Plasma Mass Spectrometry (X series ICP-MS, Thermo, Waltham, MA, USA). The resulting detection limits (in mg kg⁻¹ DW) were as follows: 0.0005, Th; 0.002, Cd, Co, Cs, U; 0.005, Cr; 0.01, As, Mn, Pb; 0.02, Cu, Sr; 0.03, Ba, Ni, Zn; 0.04, Fe; 0.1, Al, Mg; 0.4, K, Na; 1, P; and 3, Ca. Fractions of spent grains were dried and treated accordingly.

Micromalting

Grains of barley (20 g DW) and the wheat cv. JB Asano from polymetallic soil as well as the wheat cv. Kavalier from non-contaminated soil were washed twice in bideionized water and placed in weighed petri dishes 90 mm in diameter. With an intermediate break of 6 h to avoid damage by anoxic conditions, the grains were steeped in 20 mL bideionized water at 16°C for 20 h to reach a net water content of 40% by total weight. Germinating grains formed roots and sprouts up to 12-20 mm tall at 14-15°C over 6 d. The softened grains with the vegetative outgrowth that accounted for 11-12% of the dry biomass were then crushed with mortar and pestle by omitting the fermentative kilning process and suspended in glass dishes with bideionized water at 1:5 (DW/v). Mashing in a water bath was done by incubation steps of 35°C for 20 min; heating to 50°C within 15 min; 50°C for 40 min; heating to 62°C within 15 min; and 62°C for 30 min.

At this stage, half the mash as the carrier of activated enzymes was separated and left untreated. The remaining half was subjected to 68°C for 1 h to promote saccharification, boiled for 15 min to dissolve remaining starch and added to

the untreated fraction. The resulting mash was incubated at 62°C for 2 h, at 68°C for 1 h, and at 80°C for 30 min for killing enzymes.

The sweet wort was separated by sieving (0.25 mm) and rinsing the remaining draff with bideionized water to reach a total yield of 1:4.0 to the grains' initial DW (w/w). Its mineral values were re-calculated for the 1:5.5 diluted pre-alcoholic worts used for 12% beers [27, 35]. Aliquots of wort samples with specific gravities of around 1.07 were dried at 100°C to determine the content in solids, representing the malt extract. The procedures were repeated three times to obtain at least two congruent results.

Fermentation of the Pre-Alcoholic worts by *Saccharomyces Cerevisiae*

Triplicate samples of the pre-alcoholic sweet wort (10 mL) were amended with 20 mg dry baker's yeast (Netto Marken-Discount AG & Co. KG, 93142 Maxhütte-Haidhof, Germany) and incubated under anoxic conditions at 20°C for 13 d with the goal to estimate consume and precipitation of HM in the presence of yeast cells [e.g., 31,36]. The final alcoholic supernatants were centrifuged at 14,000 g for 5 min by using centrifuged sweet wort samples as references.

Statistical Treatments

SPSS 8.0 software (SPSS Inc., Chicago, IL, USA) was used to calculate the standard deviations (SD) of duplicate to quadruplicate results and linear correlations, and to perform one-way analyses of variance.

Results

Delay in Grain Germination by Heavy Metals

The initial low-temperature germination of water-steeped grains [27, 35] was significantly affected by their heavy metal load. Grains of barley and JB Asano obtained from the polymetallic HM hot spot (Tables 1 and 2) missed the critical

phytotoxicity thresholds of 5-10 mg kg⁻¹ in Cd and persisted in the lower range of the 150-220 mg kg⁻¹ in Zn [37,38]. Nevertheless, root and shoot emergence lagged 2 to 3 days behind that of gains from less contaminated soils (Figure 1). Germination was terminated when the sprouts had come to 11-12% of the original grains' DW.

Micromalting with Heavy Metal Contaminated Grains

Following micromalting, the DW of the sweet wort or malt extract per kg of dry barley grains obtained from polymetallic soil reached 51%, leaving 39% to the draff

fraction (Table 2). The wheat cv. Kavalier and the contaminated cv. JB Asano yielded 55 and 56% dry wort beside 38 and 40% draff, respectively. The permissible contamination limits of As, Cd, and Pb for fresh food grains were only violated by barley and wheat grains from the polymetallic soil in regard of Cd (Table 2). Their trace metals Cd, Cr, Cu, Mn, Pb, U, and Zn were removed to 74.3 ± 10.3% and 82.6 ± 5.5%, respectively, followed by As and Ni with around 34%, with the spent grain or draff fractions from the mash. Contemporarily, the Cd load of the dry malt extracts as the solids of the sweet worts dropped to values below the permitted 1 mg kg⁻¹ FW for food supplements (Table 2) [26].



Figure 1: Germination at 22°C of three grain sets each from winter barley (above, after 57 h) and winter wheat cv. JB Asano (below, after 100 h) that differ in their heavy metal load. Cadmium/zinc content (mg kg⁻¹ DW) of grain sets arranged left-above-right: Barley, 0.455/160-0.010/31-0.215/89; wheat, 1.55/138-0.075/27-0.820/72. Further HM with phytotoxicity potential such as As, Cu, Ni, and Pb persisted in the lower range of normal plant concentrations (Table 2).

Item	Cu	Mn	Ni	Zn	As	Cd	Cr	Pb	U	Product ^a
Winter wheat cv. Kavalier form non-contaminated soil										
Grains	4.45 ^b	26.8	0.071	30.7	0.019	0.054	0.005 ^b	0.013 ^b	0.003	-
Malt	1.38	4.46	0.126	8.74	0.03	0.015	0.022	0.026	0.004	0.5486
Draff	7.4	52.9	0.114	74.2	0.025	0.166	0.078	0.137	0.004	0.3834
Wort	0.138	0.445	0.013	0.876	0.003	0.0015	0.002	0.003	0.001	-
Winter barley from contaminated soil A										
Grains	6.1	12.5	0.37	160	0.125	0.455	0.165	0.044	0.007	-
Malt	3.85	5.08	0.644	75.5	0.146	0.202	0.028	0.049	0.008	0.5106
Draff	12.8	23	0.415	331	0.1	1.13	0.345	0.128	0.013	0.3891
Wort	0.358	0.472	0.06	7.01	0.014	0.019	0.003	0.005	0.001	-
Beer	0.051 ^c	0.481	0.06	4.82 ^c	0.001 ^c	0.004 ^c	0.004	0.004	0.001	-
Winter wheat JB Asano from contaminated soil A										
Grains	6.43	19.9	2.09	138	0.1	1.55	0.075	0.044	0.006	-
Malt	2.57	4.96	2.65	37.3	0.108	0.398	0.033	0.022	0.004	0.562
Draff	14.6	45.4	1.81	353	0.085	3.63	0.245	0.164	0.013	0.3969
Wort	0.262	0.507	0.27	3.81	0.011	0.041	0.003	0.002	0.001	-
Beer	0.030 ^c	0.458 ^c	0.239 ^c	1.45 ^c	0.002 ^c	0.001 ^c	0.005	0.004	0	-
Usual heavy metal range of herbs (in mg kg⁻¹ DW)^d										
-	2-20	14-30	0.1-3	10-100	0.01-1	0.05-0.4	0.1-1	0.1-6	<0.015	-
WHO 2011 limits for drinking water										
WHO	2	-	0.07	-	0.01	0.003	0.05	0.01	0.03	-
^a Dry weight (kg) of wort and draff recovered from one kg of dry grains. ^b Unlike the majority of values, the marked ones differ significantly at $p \leq 0.05$ from the sum of the total mineral portions represented by dry malt extract and draff. ^c Mineral concentrations in beer differ significantly at $p \leq 0.05$ from those of sweet wort. ^d Normal plant heavy metal concentrations [39,40]. Permissible contamination limits for food grains in mg kg ⁻¹ FW: As, 0.5; Pb, 0.2; Cd, 0.1 for cereals, 0.2 for wheat and rice [26].										

Table 2: Micromalting 20-g samples of barley and wheat grains (n = 2): Mean trace metal concentrations of sweet wort (1:5.5 w/v) and beer given in mg L⁻¹ and those from fresh grains, the dry malt extract (malt), and the draff fraction in mg kg⁻¹ DW.

Further trace mineral concentrations refer to the sweet worts diluted to 5.5 L per kg of dry grains as immediately used for 12% beers [27], and for their alcoholic beer state fermented by HM fishing baker's yeast [31,36].

Incubation of the sweet turbid worts with *Saccharomyces cerevisiae* formed the transparent alcoholic beer fractions and sent As (82-93%), Cd (79-98%), Cu (86-88%), and Zn (31-62%) to the trubs. Their DW came to 0.77% and 0.71% (w/v, referred to wort dilution 1:5.5) in barley and wheat beer, respectively. Thereby, Cr, Mn, Ni, Pb, and U were not removed from solution. Centrifuging wort aliquots revealed that the fermenting yeast incorporated 1.3-1.7 mg L⁻¹ of Zn as cofactor of alcohol dehydrogenase [27], but no significant amounts of the other elements. All trace mineral concentrations of wheat cv. Kavalier grains from non-contaminated soils as well as their fermenting products fitted the legislative ranges of food safety [26].

Discussion

The inherently out regulated and low metallome of seeds ensures the adequate mineral supply of the early germling by avoiding metal stress [41]. Accordingly, in spite of elevated burdens in As, Cd, Cu, Mn, U, and Zn of the polymetallic soil, barley and wheat grains persisted the widest in the usual and metal stress avoiding HM range of herbs but were somewhat higher in Cd and Zn (Table 2). Contaminations with Cd [42, 43] more than those with Cu > Zn > Pb [44] generally impair the germination of wheat and should contribute to the 2-3 days of delay in the sprouting of steeped barley and wheat grains from the polymetallic soil (Figure 1). The resulting green malt was neither kilned nor stripped of the HM enriched sprouts of 11-12% by DW as is done in the production of dry malt grains [27, 35]. Outgrowing roots and cotyledons can surpass the remaining grains in the concentrations of Ni (2x), Zn (3x), As, Cu, Hg, Pb (3-5x), and Cd and Cr (6x) [29]. The content of the contaminated and stripped barley and wheat grains would thereby drop to 71% in Zn and 55% in Cd, respectively, prior to mashing. By including the HM load of the outgrown sprouts, both draff of barley and wheat from the polymetallic

soil would concur with the tolerable limits of livestock animals in Zn (300-1000 mg kg⁻¹) [45] but not in Cd (1 mg kg⁻¹ FW) [25] (Table 2). Therefore, the spent grains should be fed, if at all, as 10% supplements.

The preferential binding of trace minerals to, and their relative enrichment in the separable draff fractions at 74-83% (Table 2) deserves attention. If surplus minerals are not located in vacuoles, they meet variable binding sites in cell walls [46, 47]. In addition, sound wheat straw [48] but also humic and fulvic acids released from senescent plants [49] and derived from biomass [50-52] unite phenolic and flavone units with terpenoids, alkanes, sugars, aliphatic and aromatic acids, tannins, heterocyclics, alcohols, amino sugars, amino acids, peptides, and proteins. Their reactive dissociable groups are mainly carboxyl (-COOH), phenolic, enolic, and alcoholic hydroxyl (-OH), methoxyl (-OCH₃), sulphhydryl (-SH), amino (-NH₂), and imino (=NH) type whose protons (H⁺) are exchangeable with metal cations [53]. Thereby, cations such as Zn²⁺ > Cu⁺ > Cu²⁺ > Ni²⁺ > Co²⁺ > Fe²⁺ > Mn²⁺ > Mg²⁺ > Ca²⁺ that most successfully compete for the respective binding sites [54] are primarily removed, with Cd having not been considered [55].

Accordingly, the remaining Cd load in the dry malt extracts from barley and wheat grains from the polymetallic soil (Cd, 39.6 mg kg⁻¹) ranged, as the fresh barley grains themselves, safely below the elevated appearing limit of 1 mg kg⁻¹ FW set for food supplements [26]. It further dropped to solid-food-grade values of ≤ 0.1 mg kg⁻¹ FW in the sweet turbid wort (Table 2) that is used at 5.5 to 6 hL per 100 kg of original grains in 12% beers [27].

The remarkably low trace mineral stock of the resulting barley and wheat beers were in the order those reported for commercial products and laid in Zn below the OIV limit for alcoholics [56-58]. Thereby, even the higher sweet wort concentrations in (As), Cr, Cu, Pb, U and in Ni from barley beer concurred with the limits set for drinking water but raised concern in regard of Cd that is confined to 0.003 mg L⁻¹ in water (Table 2) [59] and to 0.005-0.01 mg L⁻¹ in liquid baby food [26]. Alcoholic beverages have been delimited to (mg L⁻¹) 0.01 in Cd, 1.0 in Cu, 0.15 in Pb, and 5.0 in Zn by the Organization of International Vine and Wine (OIV) [60]. Zinc concentrations in this order are thereby linked with metallic taste.

Fermenting the sweet worts from barley and wheat, respectively, removed As, Cd, and Cu to 79-98%, Zn to 31-62%, but none of the traces Cr, Mn, Ni, Pb, and U from solution and led to beer concentrations in the range of drinking water (Table 2). This suggests preferential binding of As, Cd, and Cu to carboxyl groups of insufficiently degraded cell wall pectins [61] as polymers of α-(1,4)-linked D-galacturonic acid [62], to dissociable COOH, OH, and NH₍₂₎ groups of amino, imino acids, and amides as moieties of proteins [55] that may envelope residual starch granules, and to cell debris fragmented by crushing the green malt. The hypothetical daily consume of 2 L sweet wort mashed from green barley and wheat malts with their HM enriched sprouts from the

polymetallic soil would contribute 6.3% and 13.7%, respectively, to the tolerable daily intake (TDI) of 0.6 mg Cd by a 75 kg individual [26, 59]. Inputs of HM from hop supplements and from installations are not considered.

The present results concur with those of Čejka et al. [29]. Brewing water of barley was enriched with As, Cd, Cr, Hg, Ni, Se, and Pb to total concentrations of 0.1-2 mg L⁻¹ in the mash. Compared with the HM input by the current barley and wheat grains into a mash of 1:4 (w/v), the enriched mash concentrations exceeded means of the (Table 2) derived data 14x in As, 43x in Cr, and 76x in Pb. They exceeded Cd and Ni concentrations 2x and 22x, respectively, in barley but matched those introduced with the grains of JB Asano. The authors obtained quality beer due to the removal of around 76-96% of the total irritants with the spent grains fraction and another 1-11% with the trub [29]. Nevertheless, the uncontested reputation of brewery and the acceptance of its produce are based on the exclusive use of highest quality raw materials [63]. Thereby, food crops elevated in agrochemicals, mycotoxins, organic contaminants, or irritants such as As, Cd, and Pb with their carcinogenic, nephrotoxic, and cardiovascular effects, respectively [59] that multiply in their mutual interaction [64] are excluded from wholesale trade prior to distribution [65]. This comprises grains from soils that do not match with the regional hygiene guidelines [e.g., 66]. They are to be rated as forage or industrial crops in responsibility of the grower [32].

Conclusions

Unlike the drastic losses in the production of seeds (60-74%) among preceding cultures of wheat, rye, and pea from a non-remediable polymetallic soil of 39.6 mg kg⁻¹ in Cd, barley experienced the least incisions in the formation of grains (37%) [8]. Thereby, Cd exclusion led to elevated grain concentrations of 1.55 mg kg⁻¹ DW in the wheat cv. JB Asano but to 0.455 mg kg⁻¹ in barley to meet the legislative limit of 1 mg kg⁻¹ FW for livestock feed and even for human food supplements. Mashing removed Cd with the spent grain fraction to 81% in barley and to 87% in wheat. Whereas the resulting dry malt extracts concurred with the hygiene standards of Cd for food supplements, the sweet worts at the final dilution 1:5.5 met the limit of 0.1 mg kg⁻¹ FW set for solid food. Cadmium concentrations dropped further to 1-4 µg L⁻¹ during alcoholic fermenting and the removal of trub below 0.01 mg L⁻¹ set for alcoholic beverages. Thereby, the association of As, Cd, Cu and some Zn, but not of Cr, Mn, Ni, Pb, and U with wort muddying colloids such as surmised pectin and protein-starch adducts with metal chelating properties should be a topic of further studies. It is recommended that the pronounced tolerance of HM contaminated soils by barley lines and their unique trait of Cd exclusion [8] should not lightly lead to crop production on intolerably contaminated soils and to undermining of soil hygiene standards, even if subsequent crop processing eliminates the majority of contaminants from the fermentable sugar fraction. Nevertheless, with its adaptability to detrimental

climate and soil conditions and its potential health benefits, production and consume of barley deserve more attention.

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